

REVIEW

Sirtuins in wound healing

Akihiro Aioi^{1,2}

¹ Septem-Soken, Osaka, Japan

² Griffith Institute for Drug Discovery, Griffith University, Nathan, Queensland, Australia

ABSTRACT

Sirtuins (SIRT) are initially recognized as NAD⁺-dependent histone deacetylase. SIRT attract attention for their role as calorie restriction-induced “longevity proteins” to be expected to extend human life span and to promote health. As advancing studies, SIRT have been recognized as cell signaling regulators which contribute to anti-inflammation, cell differentiation and so on. Therefore, SIRT are supposed to affect wound healing which is comprised highly orchestrated complex four phases: hemostasis, inflammation, tissue formation and tissue remodeling. This review highlights the roles of SIRT in wound healing process and provides a foundation and impetus for future basic and clinical research.

Keywords: sirtuin; wound healing; anti-inflammation; re-epithelialization

ARTICLE INFO

Received: September 5, 2019

Accepted: October 11, 2019

Available online: October 22, 2019

*CORRESPONDING AUTHOR

Akihiro Aioi, Septem-Soken, Osaka,
Osaka 530-0003, Japan; a-aioi@
septem-so.com

CITATION

Aioi A. Sirtuins in wound healing.
Trends Immunother 2019; 3(2): 89–
95. doi: 10.24294/ti.v3.i2.122.

COPYRIGHT

Copyright © 2019 by author(s) and
EnPress Publisher LLC. This work is
licensed under the Creative Commons
Attribution-NonCommercial 4.0
International License (CC BY-NC 4.0).
[http://creativecommons.org/licenses/
by/4.0/](http://creativecommons.org/licenses/by/4.0/)

Introduction

Since the discovery of *silent information regulator 2* (*SIR2*) gene in 1997, *SIRT*, the *SIR2*-like genes, have been found in bacteria, plants and animals, and their function has been investigated the key roles. Sir2 protein was initially identified as a member of NAD⁺-dependent deacetylases and ADP-ribosyltransferases in *Saccharomyces cerevisiae*^[1] and subsequent studies showed that *SIR2* extends replicate life span of yeast by suppressing rDNA recombination and decreasing extrachromosomal rDNA circle^[2,3]. Following the studies on longevity of life span in *Caenorhabditis elegans* and *Drosophila*^[4–6], the studies on mammalian sirtuins embarked. Human sirtuins comprise seven members of protein (*SIRT1–7*) localized in cytosol, nucleus and mitochondria, which are involved in pleiotropic cellular functions by deacetylation of histone and/or non-histone proteins (**Table 1**). The SIRT attract attention for their role as calorie restriction-induced “longevity proteins” to be expected to extend human life span and to promote health^[7–9]. The primary function of skin, dominantly comprised by fibroblasts and keratinocytes, is to serve as a protective barrier against environment. Wound, which disrupts the primary function of skin, may lead to major disability or even death. Chronic skin ulcers such as bed sore and diabetic foot ulcer emerged as the issue to be addressed in “aged society”, as well as acute wounds caused by injury and burns in all generations. On the other hand, previous studies suggested that SIRT expressed in fibroblast and keratinocyte may concern cutaneous physiology. However, their role is gaining interest in the field of dermatology. Herein this review highlights the role of SIRT in wound healing.

Acute Wounds

Wound healing is a physiological response to restore skin integrity and is comprised highly orchestrated complex four phases: hemostasis, inflammation, tissue formation and tissue remodeling^[10]. In inflammation phase to begin with the formation of a hemostatic plug by aggregated platelets, many kinds of mediators are involved through the wound healing processes. Previously, a great deal of studies focused on growth factors, cytokines and chemokines in the process (**Table 2**)^[11]. Platelets aggregated around the wound site not only

Table 1. Sirtuins

Sirtuin	HDAC Class	Localization	Function
Sirtuin1	Class I	Nucleus Cytosol	Cell survival Life span regulation Metabolism regulation Inflammation Oxidative stress response
Sirtuin2	Class I	Nucleus Cytosol	Cell cycle regulation Nerve system development
Sirtuin3	Class I	Nucleus Cytosol Mitochondria	Mitochondrial metabolism
Sirtuin4	Class II	Mitochondria	Mitochondrial metabolism
Sirtuin5	Class III	Mitochondria	Apoptosis
Sirtuin6	Class IV	Nucleus	Genome stability DNA repair
Sirtuin7	Class IV	Nucleus	rRNA transcription regulation Cell cycle regulation

facilitate the formation of a hemostatic plug, but also secrete kinds of mediators such as platelet-derived growth factor (PDGF), that attracts and activates macrophages and fibroblasts. The activation of infiltrated cells and residing cells in wound site leads to up-regulation of proinflammatory cytokines such as interleukin (IL)-1, IL-6 and tumor necrosis factor (TNF)- α in inflammation phase. IL-1 released from macrophages and keratinocytes induces keratinocyte migration for re-epithelialization and the secretion of FGF-7 from activated fibroblasts for extracellular matrix formation^[28,29]. IL-6 expression is induced in neutrophils and macrophages immediately after wounding and the expression is sustained during healing process^[18,19]. IL-6 attracts neutrophils which cleanse the wound site of foreign particles and bacteria. Additionally, IL-6 has a mitogenic and proliferative effect on keratinocytes for re-epithelialization^[22,23]. A previous study suggested that TNF- α accelerates re-epithelialization *via* induction of FGF-7 in fibroblasts like IL-1^[31]. However, TNF- α has also been reported to suppress the wound healing *via* the induction of type V collagenase at the high concentration^[32].

Nuclear factor kappa B (NF- κ B), a transcription factor, is well-known as a major regulator of proinflammatory cytokine expression. In unstimulated cells, NF- κ B is bound to inhibitory protein, I κ B, localizing in cytosol. Extracellular stimuli recognized by receptors initiates a signaling cascade leading to the activation of I κ B kinase (IKK). The phosphorylation of I κ B by IKK induces degradation

of I κ B by proteasome and releases NF- κ B from the complex. The freed NF- κ B translocates into the nucleus where it binds to the target gene promoter region and activates gene transcription^[33]. Yeung *et al.* demonstrated that SIRT1 inhibits the transcriptional activity of NF- κ B by deacetylation of RelA/p65 subunit of NF- κ B^[34]. Indeed, resveratrol (RSV), a well-known SIRT activator, inhibited IL-6 production from normal human dermal fibroblast by lipopolysaccharide, which binds to TLR4 and activates NF- κ B, in our laboratory (**Figure 1**). TNF- α , which is a pivotal cytokine in inflammation, up-regulates the expression of matrix metalloproteinase-9 (MMP-9), IL-1 β and IL-6 in 3T3 fibroblasts. SIRT1 activation by RSV suppressed TNF- α expression by the inactivation of NF- κ B, followed by down-regulation of MMP-9, IL1 β and IL-6^[35].

Other than peptide mediators, nitric oxide (NO), a short-lived free radical, has been reported as a regulator in wound healing. NO is formed from arginine by NO synthase (NOS) which exist in three distinct isoforms, two constitutive (endothelial and neuronal) isoforms and one inducible isoform. The highest NOS activity was detected in the early phase in wound healing^[36] followed by sustained NO synthesis^[37] and the highest expression of inducible NOS was confirmed in the early phase as well^[38,39]. During the healing process, NO is involved in re-epithelialization, neovascularization and collagen synthesis^[40]. SIRT1 activation suppressed inducible NOS (iNOS) expression through the inhibition of

Table 2. Growth factors and cytokines in wound healing

Factors	Source	Function
Platele-derived growth factor ^[11]	Platelet Macrophage Keratinocyte	Fibroblast proliferation Chemoattraction
Vascular endothelial growth factor ^[11]	Epidermal cell Macrophage	Angiogenesis Increase vascular permeability
Epidermal cell growth factor ^[11,12]	Platelet Macrophage Fibroblast	Cell migration Cell proliferation
Fibroblast growth factor ^[11,13,14]	Macrophage Mast cell Endothelial cells Keratinocyte Fibroblast	Angiogenesis Fibroblast proliferation Keratinocyte migration
Transforming growth factor b1 ^[11,15,16]	Platelet Macrophage Lymphocyte Keratinocyte Fibroblast	Cell migration Chemoattraction Granulation tissue formation Re-epithelialization Extracellular matrix synthesis
Tumor necrosis factor- α ^[11,17-18]	Neutrophil Macrophage	Growth factor expression Re-epithelialization
Interleukin-1 ^[18-20]	Neutrophil Macrophage Fibroblast	Initiation of inflammation Re-epithelialization
Interleukin-6 ^[11,18,19,21-23]	Neutrophil Macrophage Fibroblast	Re-epithelialization Formation of granulation tissue Angiogenesis
Interleukin-8 ^[24-27]	Neutrophil Macrophage	Re-epithelialization

TNF- α expression^[35], SIRT1/2 has been described to enhance endothelial NOS (eNOS) activity eliciting significant increase in NO production by deacetylation at lysines 496 and 508 in the calmodulin-binding domain of eNOS^[41,42]. On the other hand, class I histone deacetylase (HDAC2), which is the only member of this class known to be S-nitrosylated directly by NO inhibits the expression of growth factors such as epidermal growth factor (EGF) by attaching the promoter regions. These suggest that post-translational S-nitrosylation of HDAC2 leads to enhance the production of growth factors by detachment of HDAC2 from the promoter regions^[43]. Consequently, SIRT-dependent NO production enhances wound closure by evocation of keratinocyte proliferation. Taken together, SIRT1 engages profoundly with the expression of proinflammatory mediators, suggesting SIRT1 could be one of the therapeutic targets for wound healing.

Chronic Wounds

Chronic wounds are defined as those which do not follow the normal healing process and show no signs of effective healing within 3 months after the injury. The cause of failure to complete wound healing is mainly to stagnate at the early inflammation phase^[44]. The features characteristic for the chronic wounds are shown in **Table 3**. Avishai *et al.* demonstrated risk factors such as autoimmune diseases, aging, obese and diabetes mellitus^[45]. In “aged-society”, the strategy of therapy for chronic wounds is the issue to be addressed, because older adults suffering from vascular disease, venous insufficiency, unrelieved pressure and diabetes mellitus, are more likely to have chronic wounds than younger people^[50]. As the morphology of resident cells is similar to that seen in senescent cells in chronic wounds, it is supposed that the treatment against cell senescence as well as against prolonged inflammation. Expression of aging

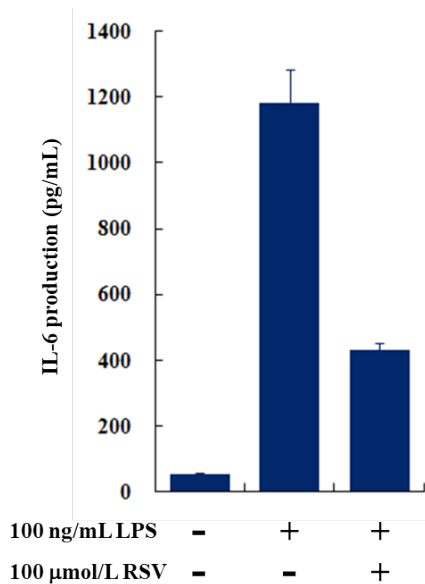


Figure 1. RSV suppressed LPS-induced IL-6 production. 100 ng/mL LPS induced abundant IL-6 production from normal human dermal fibroblasts. Addition of 100 mmol/L RSV reduced significantly the production to 38.4%.

biomarkers including procollagen I and VII, SIRT1 and SIRT6 were down-regulated in passaged human dermal fibroblasts^[51]. Growth of chronic wound fibroblasts was significantly decreased compared with fibroblasts isolated from acute wound and normal dermis^[52].

Table 3. Features in chronic wounds

Features	References
Prolonged inflammation	[44]
Excessive inflammation	[44]
Excessive neutrophil infiltration	[46]
Infection	[47]
Atypical biofilms	[48, 49]

Moreover, the abnormalities in mitogen activating protein kinase (MAPK) and Smad pathway were observed in venous ulcer fibroblasts suppressed TGF- β type II receptor expression^[53]. These suggest that TGF- β -induced collagen synthesis is suppressed in fibroblasts isolated from chronic wounds as well as senescent fibroblasts. Collagen remodeling during the transition from granulation tissue to scar is dependent on synthesis and catabolism. Collagen degradation is controlled by MMPs. Cigarette smoke exposure reduced SIRT1 expression and activity in lung tissue of A/J mice, accompanied with elevation of MMP-9. The elevation was blocked

by SRT2127, a small molecular SIRT1 activator^[54]. In skin, RSV and metformin significantly inhibited up-regulation of MMP-9 expression and prevented collagen degradation after ultra-violet irradiation^[55]. Similarly, SIRT6 has been reported to regulate negatively the expression of MMP-9^[56,57]. It is easily presumed that this negative regulation of MMP-9 expression is due to inhibition of NF- κ B pathway, because MMP-9 expression is regulated by NF- κ B which is a deacetylation target of SIRT6. Like so, suppressed MMP-9 expression contributes to collagen re-modeling in dermis. However, MMP-9 deficiency leads to impaired wound healing, because MMP-9 also contributes keratinocyte migration in re-epithelialization phase. On the other hand, high glucose impaired keratinocyte migration by inducing levels of MMP-9 expression in diabetic mouse model. This glucose-sensitive elevation of MMP-9 expression was blocked by deletion of FOXO1, which is also a deacetylation target of SIRT1, concomitant with improved wound healing^[58]. Therefore, the appropriate expression of MMP-9 is required for orchestrated wound healing process.

Conclusion

SIRT6 attract attention for their role as calorie restriction-induced “longevity proteins” with expectation to extend human life span by promoting health and wellness. As studies advance on SIRT6, it has been emerged that SIRT6 are involved in pleiotropic functions *via* deacetylation of histone and/or non-histone proteins (**Figure 2**)^[59]. Of them, inflammation, cell proliferation and cell migration are vital events in wound healing process. SIRT6 have not only anti-inflammatory effects but also promotive effect on cell proliferation and cell migration to enhance wound healing, suggesting SIRT6 activation could be one of the therapeutic strategies for wound healing. SIRT6 activators have been found in nature and synthesized such as RSV and its derivatives^[60]. The effect of RSV on wound healing and chronic diseases has been evaluated not only in animal model and human EpiDerm full thickness model, but also in clinical applications^[61-63]. Taken together, SIRT6 are involved in orchestrated wound healing processes and its activation provides an approach for acceleration of wound therapy. RSV could be a major candidate compound for wound healing because it is a polyphenol found in nature such as within plants, or the consumed such as grapes and wine.

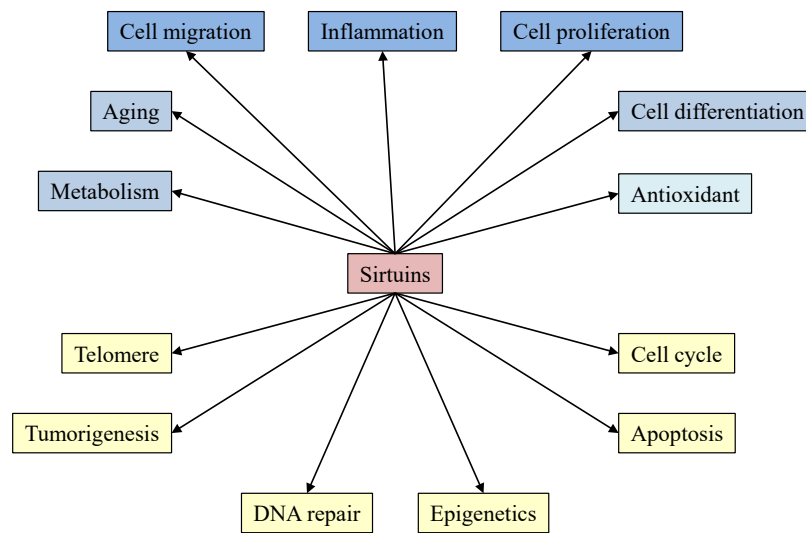


Figure 2. Multi-functions of SIRT. SIRTs possess multi-functions other than originally recognized function of life longevity. Blue-shaded functions are vastly involved in wound healing.

Conflict of interest

The author declares no potential conflict of interest with respect to the research, authorship, and/or publication of his article.

References

1. Imai S, Armstrong CM, Kaerberlein M, *et al.* Transcriptional silencing and longevity protein Sir2 is an NAD-dependent histone deacetylase. *Nature* 2000; 403(6771): 795–800. doi: 10.1038/35001622.
2. Sinclair DA, Guarente L. Extrachromosomal rDNA circles—A cause of aging in yeast. *Cell* 1997; 91(7): 1033–1042. doi: 10.1016/S0092-8674(00)80493-6.
3. Kaerberlein M, McVey M, Guarente L. The SIR2/3/4 complex and SIR2 alone promote longevity in *Saccharomyces cerevisiae* by two different mechanisms. *Genes Dev* 1999; 13(19): 2570–2580. doi: 10.1101/gad.13.19.2570.
4. Tissenbaum HA, Guarente L. Increased dosage of a *sir-2* gene extends lifespan in *Caenorhabditis elegans*. *Nature* 2001; 410(6825): 227–30. doi: 10.1038/35065638.
5. Wang Y, Tissenbaum HA. Overlapping and distinct functions for a *Caenorhabditis elegans* SIR2 and DAF-16/FOXO. *Mech Ageing Dev* 2006; 127(1): 48–56. doi: 10.1016/j.mad.2005.09.005.
6. Rogina B, Helfand SL. Sir2 mediates longevity in the fly through a pathway related to calorie restriction. *Proc Natl Acad Sci USA* 2004; 101(45): 15998–16003. doi: 10.1073/pnas.0404184101.
7. Colman R, Anderson R, Johnson S, *et al.* Caloric restriction delays disease onset and mortality in rhesus monkey. *Science* 2009; 325(5937): 201–204. doi: 10.1126/science.1173635.
8. Guarente L, Picard F. Calorie restriction—The SIR2 connection. *Cell* 2005; 120(4): 473–482. doi: 10.1016/j.cell.2005.01.029.
9. Haigis M, Mostoslavsky R, Haigis K, *et al.* SIRT4 inhibits glutamate dehydrogenase and opposes the effects of calorie restriction in pancreatic beta cells. *Cell* 2006; 126(5): 941–954. doi: 10.1016/j.cell.2006.06.057.
10. Singer AJ, Clark RAF. Cutaneous wound healing. *N Eng J Med* 1999; 341(10): 738–746. doi: 10.1056/NEJM199909023411006.
11. Barrientos S, Stojadinovic O, Golinko MS, *et al.* Growth factors and cytokines in wound. *Wound Rep Reg* 2008; 16(5): 585–601. doi: 10.1111/j.1524-475X.2008.00410.x.
12. Shen C, Sun L, Zhu N, *et al.* Lindlin-1 contributes to EGF-induced re-epithelialization in skin wound healing. *Int J Mol Med* 2017; 39(4): 949–959. doi: 10.3892/ijmm.2017.2911.
13. Meyer M, Muller AK, Yang J, *et al.* FGF receptors 1 and 2 are key regulators of keratinocyte migration *in vitro* and in wounded skin. *J Cell Sci* 2012; 125(Pt 23): 5690–5701. doi: 10.1242/jcs.108167.
14. Takamiya M, Saigusa K, Nakayashiki N, *et al.* Studies on mRNA expression of basic fibroblast growth factor in wound healing for wound age determination. *Int J Legal Med* 2003; 117(1): 46–50. doi: 10.1007/s00414-002-0354-3.
15. Zhang C, Tan CK, McFarlane C, *et al.* Myostatin-null mice exhibit delay skin wound healing through the blockade of transforming growth factor- β signaling by decorin. *Am J Cell Physiol* 2012; 302(8): C1213–C1225. doi: 10.1152/ajpcell.00179.2011.

16. Ishida Y, Kondo T, Takayasu T, *et al.* The essential involvement of cross-talk between IFN- γ and TGF- β in the skin wound-healing process. *J Immunol* 2004; 172(3): 1848–1855. doi: 10.4049/jimmunol.172.3.1848.
17. Ashcroft GS, Jeoung MJ, Ashworth JJ, *et al.* Tumor necrosis factor-alpha (TNF- α) is a therapeutic target for impaired cutaneous wound healing. *Wound Rep Reg* 2012; 20(1): 38–49. doi: 10.1111/j.1524-475X.2011.00748.x.
18. Grellner W. Time-dependent immunohistochemical detection of proinflammatory cytokines (IL-1 β , IL-6, TNF- α) in human skin wounds. *Forensic Sci Int* 2002; 130(2–3): 90–96. doi: 10.1016/S0379-0738(02)00342-0.
19. Grellner W, George T, Wilske J. Quantitative analysis of proinflammatory cytokines (IL-1 β , IL-6, TNF- α) in human skin wounds. *Forensic Sci Int* 2000; 113(1–3): 251–264. doi: 10.1016/S0379-0738(00)00218-8.
20. Ishida Y, Kondo T, Kimura A, *et al.* Absence of IL-1receptor antagonist impaired wound healing along with aberrant NF- κ B activation and a reciprocal suppression of TGF- β signal pathway. *J Immunol* 2006; 176(9): 5598–5606. doi: 10.4049/jimmunol.176.9.5598.
21. Gallucci RM, Sugawara T, Yucesoy B, *et al.* Interleukin-6 treatment augments cutaneous wound healing in immunosuppressed mice. *J Interferon Cytokine Res* 2001; 21(8): 603–609. doi: 10.1089/10799900152547867.
22. Gallucci RM, Sloan DK, Heck JM, *et al.* Interleukin 6 indirectly induces keratinocyte migration. *J Invest Dermatol* 2004; 122(3): 764–772. doi: 10.1111/j.0022-202X.2004.22323.x.
23. McFarland-Mancini MM, Funk HM, Pluch AM, *et al.* Differences in wound healing in mice with deficiency of IL-6 versus IL-6 receptor. *J Immunol* 2010; 184(12): 7219–7228. doi: 10.4049/jimmunol.0901929.
24. Kondo T, Oshima T, Mori R, *et al.* Immunohistochemical detection of chemokines in human skin wounds and its application to wound age determination. *Int J Legal Med* 2002; 116(2): 87–91. doi: 10.1007/s004140100260.
25. Engelhardt E, Toksoy A, Goebeler M, *et al.* Chemokines IL-8, GRO α , MCP-1, IP-10, and Mig are sequentially and differentially expressed during phase-specific infiltration of leukocyte subsets in human wound-healing. *Am J Pathol* 1998; 153(6): 1849–1860. doi: 10.1016/S0002-9440(10)65699-4.
26. Ishida Y, Gao JL, Murphy PM. Chemokine receptor CX3CR1 mediates skin wound healing by promoting macrophage and fibroblast accumulation and function. *J Immunol* 2008; 180(1): 569–579. doi: 10.4049/jimmunol.180.1.569.
27. Aibina JE, Mills CD, Henry WL, *et al.* Temporal expression of different pathways of l-arginine metabolism in healing wounds. *J Immunol* 1990; 144(10): 3877–3880.
28. Komine M, Rao LS, Kaneko T, *et al.* Inflammatory versus proliferative process in epidermis. Tumor necrosis factor- α induces K6b keratin synthesis through a transcriptional complex containing NF κ B and C/EBP β . *J Biol Chem* 2000; 275(41): 32077–32088. doi: 10.1074/jbc.M001253200.
29. Tang A, Gilchrist BA. Regulation of keratinocyte growth factor gene expression in human skin fibroblasts. *J Dermatol Sci* 1996; 11(1): 41–50. doi: 10.1016/0923-1811(95)00418-1.
30. Sato M, Sawamura D, Ina S, *et al.* *In vivo* introduction of the *inteleukin-6* gene into human keratinocyte: Induction of epidermal proliferation by the fully spliced form of interleukin-6, but not by the alternatively spliced form. *Arch Dermatol Res* 1999; 291(7–8): 400–404. doi: 10.1007/s004030050429.
31. Brauchle M, Angermeyer K, Hubner G, *et al.* Large induction of keratinocyte growth factor expression by serum growth factors and proinflammatory cytokines in cultured fibroblasts. *Oncogene* 1994; 9(11): 3199–3204.
32. Unemori EN, Hibbs MS, Amento EP. Constitutive expression of 92-kD gelatinase (type V collagenase) by rheumatoid synovial fibroblasts and its induction in normal human fibroblasts by inflammatory cytokines. *J Clin Invest* 1991; 88(5): 1656–1662. doi: 10.1172/JCI115480.
33. Napetschnig J, Wu H. Molecular basis of NF- κ B signaling. *Annu Rev Biophys* 2013; 42: 443–468. doi: 10.1146/annurev-biophys-083012-130338.
34. Yeung F, Hoberg JE, Ramsey CS, *et al.* Modulation of NF- κ B-dependent transcription and cell survival by the SIRT1 deacetylase. *EMBO J* 2004; 23(12): 2369–2380. doi: 10.1038/sj.emboj.7600244.
35. Zhu X, Liu Q, Wang M, *et al.* Activation of Sirt1 by resveratrol inhibit TNF- α induced inflammation in fibroblasts. *PloS One* 2011; 6(11): e27081. doi: 10.1371/journal.pone.0027081.
36. Schaffer MR, Tantry U, Barbul A, *et al.* Nitric oxide metabolism in wounds. *J Surg Res* 1997; 71(1): 25–31. doi: 10.1006/jsre.1997.5137.
37. Frank S, Madlener M, Pfeilschiter J, *et al.* Induction of inducible nitric oxide synthase and its corresponding tetrahydrobiopterin-cofactor-synthesizing enzyme GTP-cyclohydrolase I during cutaneous wound healing. *J Invest Dermatol* 1998; 111(6): 1058–1064. doi: 10.1046/j.1523-1747.1998.00434.x.
38. Reichner JS, Meszaros AJ, Louis CA, *et al.* Molecular and metabolic evidence for the restricted expression of inducible nitric oxide synthase in healing wounds. *Am J Pathol* 1999; 154(4): 1097–1104. doi: 10.1016/S0002-9440(10)65362-X.
39. Paulsen SM, Wurster SH, Nanney LB. Expression of inducible nitric oxide synthase in human burn wounds. *Wound Rep Reg* 1998; 6(2): 142–148. doi: 10.1046/j.1524-475X.1998.60208.x.
40. Rizk M, Witte MB, Barbul A. Nitric oxide and wound healing. *World J Surg* 2004; 28(3): 301–306. doi: 10.1007/s00268-003-7396-7.

41. Mattagajasingh I, Kim CK, Naqvi A, *et al.* SIRT promotes endothelium-dependent vascular relaxation by activating endothelial nitric oxide synthase. *Proc Natl Acad Sci* 2007; 104(37): 14855–14860. doi: 10.1073/pnas.0704329104.
42. Valente S, Mellini P, Spallotta F, *et al.* 1,4-Dihydropyridines active on the SIRT1/AMPK pathway ameliorate skin repair and mitochondrial function and exhibit inhibition of proliferation in cancer cells. *J Med Chem* 2016; 59(4): 1471–1491. doi: 10.1021/acs.jmedchem.5b01117.
43. Spallotta F, Cencioni C, Straino S, *et al.* A nitric oxide-dependent crosstalk between class I and II histone deacetylases accelerates skin repair. *J Biol Chem* 2013; 288(16): 11004–11012. doi: 10.1074/jbc.M112.441816.
44. Eming SA, Krieg T, Davidson JM. Inflammation in wound repair: Molecular and cellular mechanisms. *J Invest Dermatol* 2007; 127(3): 514–525. doi: 10.1038/sj.jid.5700701.
45. Avishai E, Yeghiazaryan K, Golubnitschaja O. Impaired wound healing: Facts and hypotheses for multi-professional considerations in predictive, preventive and personalized medicine. *EPMA J* 2017; 8(1): 23–33. doi: 10.1007/s13167-017-0081-y.
46. Martin P, Nunan R. Cellular and molecular mechanisms of repair in acute and chronic wound healing. *Br J Dermatol* 2015; 173(2): 370–378. doi: 10.1111/bjd.13954.
47. Edwards R, Harding KG. Bacteria and wound healing. *Curr Opin Infect Dis* 2004; 17(2): 91–96. doi: 10.1097/00001432-200404000-00004.
48. Wolcott RD, Rhoads DD, Dowd SE. Biofilm and chronic wound inflammation. *J Wound Care* 2008; 17(8): 333–341. doi: 10.12968/jowc.2008.17.8.30796.
49. Thomson CH. Biofilms: Do they affect wound healing? *Int Wound J* 2011; 8(1): 63–67. doi: 10.1111/j.1742-481X.2010.00749.x.
50. Gould L, Abadir P, Brem H, *et al.* Chronic wound repair and healing in older adults: Current status and future research. *J Am Geriatr Soc* 2015; 63(3): 427–438. doi: 10.1111/jgs.13332.
51. Kim KS, Park HK, Lee JW, *et al.* Investigate correlation between mechanical property and aging biomarker in passaged human dermal fibroblasts. *Microsc Rec Tech* 2015; 78(4): 277–282. doi: 10.1002/jemt.22472.
52. Agren MS, Steenfors HH, Dabelsteen S, *et al.* Proliferation and mitogenic response to PDGF-BB of fibroblasts isolated from chronic venous leg ulcers is ulcer-age dependent. *J Invest Dermatol* 1999; 112(4): 463–469. doi: 10.1046/j.1523-1747.1999.00549.x.
53. Kim BC, Kim HT, Park SH, *et al.* Fibroblasts from chronic wounds show altered TGF- β -signaling and decreased TGF- β type II receptor expression. *J Cell Physiol* 2003; 195(3): 331–336. doi: 10.1002/jcp.10301.
54. Nakamura Y, Vuppusetty C, Wada H, *et al.* A protein deacetylase is a negative regulator of metalloproteinase-9. *FASEB J* 2009; 23(9): 2810–2819. doi: 10.1096/fj.08-125468.
55. Lee JS, Park KY, Min HG, *et al.* Negative regulation of stress-induced matrix metalloproteinase-9 by Sirt1 in skin tissue. *Exp Dermatol* 2010; 19(12): 1060–1066. doi: 10.1111/j.1600-0625.2010.01129.x.
56. Kang L, Hu J, Weng Y, *et al.* Sirtuin 6 prevents matrix degradation through inhibition of the NF- κ B pathway intervertebral disc degeneration. *Exp Cell Res* 2017; 352(2): 322–332. doi: 10.1016/j.yexcr.2017.02.023.
57. Thandavaryan RA, Garikipati VNS, Joladarashi D, *et al.* Sirtuin-6 deficiency exacerbates diabetes-induced impairment of wound healing. *Exp Dermatol* 2015; 24(10): 773–778. doi: 10.1111/exd.12762.
58. Zhang C, Lim J, Jeon HH, *et al.* FOXO1 deletion in keratinocytes improves diabetic wound healing through MMP9 regulation. *Sci Rep* 2017; 7(1): 10565. doi: 10.1038/s41598-017-10999-3.
59. Serravallo M, Jagdeo J, Glick SA, *et al.* Sirtuins in dermatology: Applications for future research and therapeutics. *Arch Dermatol Res* 2013; 305(4): 269–282. doi: 10.1007/s00403-013-1320-2.
60. Villalba JM, Alcain FJ. Sirtuin activators and inhibitors. *Biofactors* 2012; 38(5): 349–359. doi: 10.1002/biof.1032.
61. Zhao P, Sui BD, Liu N, *et al.* Anti-aging pharmacology in cutaneous wound healing: Effects of metformin, resveratrol, and rapamycin by local application. *Aging Cell* 2017; 16(5): 1083–1093. doi: 10.1111/accel.12635.
62. Lephart E, Sommerfeldt JM, Andrus MB. Resveratrol: Influences on gene expression in human skin. *J Func Foods* 2014; 10: 377–384. doi: 10.1016/j.jff.2014.07.017.
63. Wahab A, Gao K, Jia C, *et al.* Significance of resveratrol in clinical management of chronic diseases. *Molecules* 2017; 22(8): 1329. doi: 10.3390/molecules22081329.