REVIEW ARTICLE

Harnessing the crosstalk of adipocytes, autophagy, and immune cells for immunotherapy in obesity

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ABSTRACT

As a self-degradative and recycling program, autophagy plays an essential role in homeostasis and life. The connection between autophagy and the status of the adipose tissue (white or beige/brown) links to metabolic diseases such as obesity, type two diabetes mellitus (T2DM). Moreover, autophagy and the renin-angiotensin physiological system play a pivotal role in metabolic syndrome, a disease that can disrupt homeostasis in different organs, including adipose tissue. The crosstalk in adipose tissue maintains low inflammation, brown adipocytes, and autophagic machinery under control. The JAK-STAT signalization pathway and the paracrine action of hormones, adipokines, and cytokines play a role in maintaining the status of low inflammation, brown adipocytes, and autophagic machinery to harness the utmost for obesity immunotherapy.

Keywords: Autophagy; Adipocytes; Immune System; Leptin; Obesity; Type Two Diabetes Mellitus (T2DM)

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1. Introduction

Obesity is a condition of excessive fat in adipose tissue. It has become a global problem of concern, especially in young adults (under 20 years old), that represent 25% to 30% of adult $people^{[1,2]}$. The lack of physical activity, overnutrition, and importantly the diet (rich in carbohydrates and fats) contribute greatly to increase this problem^[3–5]. The problem have become ruthless and hard to overcome because those with obesity are predisposed to suffer from other chronic complications, i.e. type 2 diabetes mellitus, hypertension, cardiovascular disease, as well as to suffer from other chronic diseases such as cancer^[6–9]. Several studies and efforts have been made to have an adequate management of obesity in different aged populations. Studies at the level of basic clinic research focused to understand the links between obesity and the implication of this condition at the cellular and organismic level. At this point, it is known but not understood clearly that there are key links of the crosstalk between the dysfunction in any of organs or tissues and insulin sensitivity/resistance^[8-11].

For example, in obesity, in addition to the surplus of nutrients and energy stored in the adipose tissue and in non-adipose tissue as fat, there are production of inflammatory cytokines, adipokines and reactive oxygen species^[12–14]. These provoke damage at tissue (liver, or skeletal muscle) or at cellular levels (adipocytes differentiation)^[14,15]. Indeed the surplus of nutrients can provoke systemic lipotoxicity due to the excessive fat accumulation, elevating thus, serum free fatty acid levels^[16]. Under these settings, the health condition of the individual is really compromised, because the potential links of the adipose tissue dysfunction with other metabolic organs and cellular processes^[9,11,17].

Adipose tissue (AT), considered a dynamic endocrine organ, key in energy and metabolic homeostasis closely related to the energy storage and to the secretion of different components and factors that participate in the regulation of several physiological functions (i.e. metabolism, reproduction coagulation and cardiovascular function). AT secretes a great amount of factors that includes those with endocrine function (adiponectina, leptin, omentin, resistine, visfatine, apeline, vaspine, apelin, various growth factors, sex steroids), and those with immune function (adipsine or factor D of the complement, hepcidine, haptoglobulin)^[18,19]. In fact, AT and the lymphoid tissue anatomically is closer to the adipose tissue, generating a microenvironment thus helping the immune system to answer^[20,21]. Other components also secreted by AT are the protein 4 ligand of retinol (RBP-4), non-esterified fatty acids (NEFA), inhibitor of the activator of plasminogen (PAI-1), 11 β-dehidroxiesteroide dehidrogenasa, prostaglandins, angiotensinogen that participate metabolic, cardiovascular in function^[17,22,23].

Adipose tissue (AT) is conformed by the bona fide cells as pre-adipocytes, adipocytes, endothelial, and immune cells. The two principal types of adipocytes are the white and brown with an intermediate state of adipocytes, and the brown adipocytes that depend mostly of the lipid and mitochondria content. Thus, beige adipocytes are with an intermediate amount of each of these components^[15]. Macrophages and eosinophils function in adipose tissue as the vin and yang innate immune cells in AT. Macrophages in obese adipose tissue increase and are associated with inflammation and metabolic disease, specifically insulin resistance^[24,25]. Together, all these cells make up AT and contribute to the cellular composition and regulation of obesity and metabolic dysfunction^[25,26].

Specifically, brown and white adipocytes (WAT/BAT) secrete several components, enzymes, hormones, and growth factors that activate the

JAK-STAT pathway^[27,28]. Thus, adipocytes in AT establish paracrine crosstalk of the immune cells and autophagy, representing a platform for immunotherapy^[29-32]. Furthermore, adipocyte differentiation is crucial for metabolic homeostasis^[13]. In AT, adipogenesis is carried out in two stages: commitment of mesenchymal stem cells to a pre-adipocyte fate and terminal differentiation, and a signalization pathway of WNT and RHO-family GTPase cascade has been involved in this. In the terminal differentiation of committed preadipocytes through the epigenomic activation of peroxisome proliferator-activator receptor-gamma (PPARA- γ) and the adipogenic stimuli^[13,15]. The adipocyte gene expression in AT is maintained through the coordination of PPAR-y and the CCAAT/enhancer binding protein (C/EBP) transcription factors^[33]. Understanding this mechanism of adipogenesis and the players involved in it, can also provide therapeutic targets against metabolic diseases^[11,33].

Since autophagy is an adaptive eukaryotic process, pivotal for cellular homeostasis, any stress or cellular, organelle damage triggers the induction of autophagy^[34,35]. Indeed, there has been a recent study on autophagic response in obesity condition and the links between insulin action and the type 2 diabetes mellitus^[29]. Thus, autophagy can be activated either under low nutrients (constitutive activity) intake or in overnutrition^[29,35]. A defect or impairment of autophagy machinery program represent a failure and a cause of disease, exacerbated inflammatory responses and organelle dysfunction^[36,37]. For example, lipids or glycogen can compromise hepatic metabolic function and affect insulin action, exacerbating insulin resistance and possibly other metabolic pathologies associated to obesity. In obesity, it has been found an impairment in autophagy and insulin signaling^[33,38].

Another potential link is that autophagy acts in concert with the immune cells present in AT, and with endocrine renin-angiotensin system (RAS system) for regulation of blood pressure and fluid balance^[33,39], dysregulation of which can exhibit abnormalities in obesity. Furthermore, autophagy and obesity connections are related to the ER stress^[40,41]. In fact, ER can provide the membranes for the autophagosome formation^[15,29,41], and obesity is characterized by ER stress^[37]. In experimental ER stress, autophagy is induced, and several canonical UPR (unfolded protein response) pathways are activated^[35,39]. ER stress can have an effect in the pancreatic beta cell function, and thereby in insulin secretion. Thus, autophagy also plays a role to keep homeostasis at the level not only of ER but of the beta cell function, critical in metabolic diseases, such as obesity and diabetes^[38]. In obesity, insulin action and mTOR, are altered, both of which are autophagy regulators^[41,42]. Therefore, dysregulation of autophagy constitutes a critical component of obesity and contribute to metabolite dysfunction.

Current studies and evidence from the literature have shed light on the close crosstalk sustained among different physiological systems to keep homeostasis and individuals functional with life. Work is ongoing in the lab to investigate and determine the status of autophagy, adipokines, and the immune system in obesity in young individuals. These might provide potential therapeutic targets for prevention and immunotherapies.

Adipose tissue and adipogenesis. The adipose tissue is a dynamic endocrine organ that functions in energy storage. It is also the primary site of inflammation in obesity and secretes several participant factors of the immune response^[27,43]. The secretion of a great variety of proteins enables it to participate importantly in the regulation of appetite, metabolism, reproduction, coagulation, and cardiovascular function^[12,13,44,45]. The deposit sites of adipose tissue can be subcutaneous (80%) or visceral (20%). The vascularized tissue has higher sympathetic innervation and many β adrenergic receptors, enabling it with higher activity and relation with the associated pathology with obesity.

In humans, there are two types of adipose tissue, brown (responsible for thermogenesis) and white adipose tissue (fat storage and secretion of molecules)^[12,13,44,45]. In the adipose tissue of obese individuals, the status is characterized by an increase in adipocytes, hypertrophy (increase in size), or hyperplasia (increase in the number). A remarkable anatomical feature is that lymphoid tissue is very close to adipose tissue, generating a microenvironment that helps the immune system^[12,13,44] (**Figure 1A**). Both types of tissues interact locally through common mediators—cytokines, adipokines (leptin, adiponectin, resisting, visfatin, apelin, Caspian adipsin), factor D of the complement, hepcidin, retinol ligand protein 4 (RBP4), non-esterified fatty acids—An inhibitor of the plasminogen (PAI-1), 11 β -di-hydroxysteroid dehydrogenase^[13,19,42] (**Figure 1B**).

Autophagy is a biological and cellular process of catabolism by recycling products of metabolism. It is a mechanism of autoregulation of renewal to eliminate intracellular components, excessive nutrients, toxic protein aggregates, damaged organelles, and invasive microorganisms^[30,46]. This process plays a pivotal role in maintaining energy homeostasis and protection against stress.

Autophagy occurs in specialized vacuoles of double membrane-denominated autophagosomes. It requires the participation of lysosomes^[46,47]. Autophagy self-program is formed by a set of associated ATG proteins that conform machinery for cytoplasm checking quality control. The status of autophagy under stress conditions, starvation, or nutrient limitation (glucose, amino acids, growth factors, oxygen, pathogens) is the upregulation of ATG genes, particularly ATG5, potentially linked with inflammation^[48,49]. Autophagy keeps low levels of homeostasis and survival in anabolic reactions^[35]. Activation of autophagy occurs under environmental stress, malnutrition, and other factors, leading to a metabolic dysfunction: at the level of adipocyte differentiation (an increase in white adipose tissue), a dysregulation in lipid metabolism (an imbalance in lipids) and activation of the JAK-STAT signalization pathways)^[36,47,50]. (Figures **1A** and **1B**)

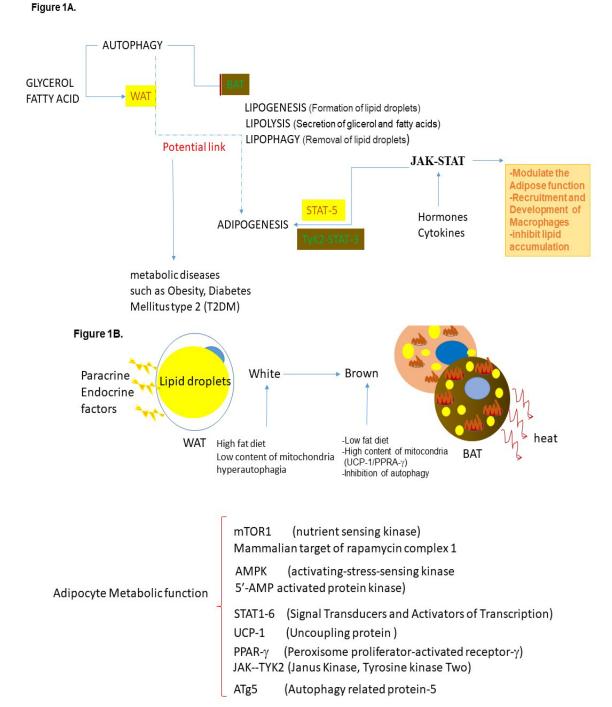


Figure 1. Schematic representation of the metabolic pathways, lipids metabolism, autophagy connected through the JAK-STAT signalization pathway. To maintain homeostasis in adipose tissue makes necessary that all the metabolic pathways function properly connected with autophagy machinery for elimination of unwanted material and cellular damaged organelles. The JAK STAT activators have the enormous responsibility of the functionality of the white and/or brown adipocytes. (**A**) The link are the paracrine secreted components of adipocytes and the immune cells that influence the state of the inflammation status and of the immunometabolic health. (**B**) Features ad hoc of the white and beige/brown adipocytes, the function and development depend of the transcriptional control of the JAK-STAT activator and of the autophagy machinery. White adipocytes are the lipid storage compartment in adipose tissue while brown adipocytes are responsible of the expenditure of the energy, to release energy under cold and beta-adrenergic receptor stimulation.

Regulation of autophagy is necessary to fit an individual metabolic profile and warrant a proper balance in adipose tissue metabolism function. Dysregulation of autophagy correlates with neurodegenerative and cardiovascular disorders. Therefore, the therapeutic implication of the modulation of autophagy constitutes with of the most exciting areas of research^[32,35,51]. When autophagy machinery is dysfunctional for a prolonged period (high fat or fructose diet), accumulation of unwanted proteins and organelles in adipose tissue, liver, muscle, and pancreas is observed. Under these settings, autophagy becomes detrimental and eventually induces metabolic dysfunction and metabolic diseases such as obesity and type 2 diabetes mellitus (T2DM) and other effects on health (cachexia, hepatic steatosis, rarosis, atherosclerosis)^[27].

2. Autophagy in obesity

Obesity constitutes a health problem, especially in developed countries. In Mexico. around 70% of Mexicans suffer from being overweight, and almost a third proportion develop obesity. Obesity is associated with the development of metabolic syndrome^[39,42], leading to diabetes and cardiovascular diseases, but also affects muscular tissue, bones, and some types of cancer^[1,2,39]. Hypercaloric foods intake (rich in fat, salt, and sugar) but deficient in minerals and vitamins can cause obesity. Other factors that lead to obesity are some physiological disorders: hypothyroidism and the Cushing disease, pharmacological drugs; anti-depressive, corticosteroids, nutrition diet, smoking habits, genetic factors, age, and race $^{[2,30,52]}$.

Adipogenesis and autophagy have been proposed as potential links to metabolic diseases such as obesity and diabetes mellitus. In referring to the role of autophagy in obesity, it has been reported that there is a combined expression of proteins associated with autophagy ATG5/LC3 (LC3-II)/p62 (ubiquitin-bonding scaffold protein) in obese individuals and mice^[53,54]. Autophagy can be involved in the browning of the white adipose tissue. It can affect the metabolism equilibrium of lipids. Furthermore, the role of autophagy in the development of obesity has been related to insulin sensibility. (**Figures 1A** and **1B**)

Autophagy can degrade the cytoplasmic lipids in hepatocytes, a process called "hypophagia"^[49]. The equilibrium related to the quantity of white adipose tissue and brown adipose tissue can affect the storage of lipids and the energetic homeostasis of the body. Mice with selective alteration of ATG7 are brown with decreased white adipose mass and are sensible to the insulin, and show an accumulation of lipids due to defects in the elimination process of lipids (lipophagia). In addition, the absence of ATG7 in preadipocytes 3T3-L1 decreased protein levels and factors of adipocyte differentiation. Similarly, the lack of ATG5 or the pharmacological inhibition of autophagy has similar effects^[57]. Knockout mice with autophagy genes in adipose tissue show defects in adipocyte differentiation, skeletal muscle, or liver, leading to leanness, obesity resistance, and induced diabetes because of the diet^[35]. Therefore, autophagy regulates the accumulation of corporal or body lipids controlling the differentiation of the adipocytes and determining the equilibrium between the white and brown fat^[19,24]. By another hand, there is a relationship between some hormones that regulate appetite (ghrelin), and autophagy.

The ghrelin activation of the pathway PI3K/Akt/Bcl-2 inhibits the activation of autophagy. This effect can be interrupted by an inhibitor of the Akt kinase^[48]. In referring to the studies approaching the role of the autophagy genes, those described in the mouse and human models were cited. In preclinic models (obese mice)^[34,57], deletion in the gene ATG7 causes a phenotype of brown adipose tissue and a high metabolic rate. These knockout mice keep weight and are resistant to obesity. Moreover, obese mice show a decreased expression of ATG5 and ATG7. However, in a clinic model using explants in visceral adipose tissue and subcutaneous tissue (WAT), expression of ATG5, IC3, and IC3B is increased^[36]. Autophagy dysregulation leads to problems in overweight, obesity, and diabetes. It predisposes to other neurodegenerative diseases as hypertension and cardiovascular diseases^[6,32]. Weight loss and autophagy regulation targeted to improve metabolic health targeting in anti-obese therapies. Adipogenesis increase when autophagy is inhibited facilitating, thus, weight loss and improving metabolic health^[6,32]. Furthermore, the role of autophagy in adipocytes differentiation can be through the activation of the autophagy by the angiotensin II pathway modulated by NADPH oxidase and ROS, both targets of cellular stress, inflammation, and cellular infiltration^[12,13]. The adipose autophagy activated regulates the increase in C/EBP, Fabp4, Agpat2, and FAS, related to the differentiation and adipose maturation. The activation of the autophagy could be through NADPH oxidase mediated by the production of angiotensin II and ROS, as well as other triggering factors like cellular stress inflammation and macrophages infiltration^[39,42].

3. The status of autophagy in obesity

Autophagy is inactive during obesity conditions because hyper nutrition inhibits AMPK (serine/threonine kinase AMP-activated protein kinase complex) and activates mTOR1 (nutrient-sensing kinase) (mammalian target of rapamycin complex 1)^[30]. In obesity, mTOR1 is positively regulated and associated with anabolic metabolism in the liver^[34]. The resistance to insulin and hyperinsulinemia is attributed to the inhibition of autophagy in obesity^[6]. In recent times, some studies have related autophagy with the regulation of lipids metabolism, and today it is known to be through four pathways. (**Figures 1A** and **1B**).

1) The metabolites released from the lipids metabolism activate the mTOR pathway decreasing the activation of autophagy^[30].

2) Changing the morphology of the lipid membrane and the transport of the vesicles of binding to effectors proteins.

3) Some lipidic molecules facilitate the modification of the proteins to regulate autophagy for example in the lipidation of the family of proteins Atg8/LC3.

4) The lipidic molecules regulate autophagy by the control in the distribution of specimens of lipids in the double lipidic membrane.

The resistance to insulin and hyperinsulinemia is associated with the inhibition of autophagy during obesity. Obese mice show a low expression of ATG5 and ATG7 and an inhibition of autophagosome biogenesis^[52,55].

Some other studies in humans are related to the increase in the protein expression and the ARNm of ATG5, IC3A, and IC3B^[36]. Moreover, the events involved in the acquired pathogenesis of the over-

weight and obesity condition at the physiological level, the body suffers a significant dysregulation in diverse processes^[36,39,56], for example, in mechanisms of recycling and destruction to the cellular level of proteins, cellular remnants or whole organelles, in senescence or anormal, as well as substances in the cytoplasm due to autophagy. This process is activated under different conditions like stress, starvation, and microbial or in the development of anormal cells^[35]. In the regulation of the metabolism of lipids, it is proposed that the autophagy machinery intact is necessary for the biogenesis of lipids.

3.1 The status of the immune system in obesity

In the last years, it has been associated obesity with a chronic inflammatory process of low intensity. Obesity alters the metabolism of the adipose tissue and endocrine functions, leading to an increase in the release of hormones, fatty acids, and pro-inflammatory molecules that contribute to the association of own complications of the disease^[58-60]. When persons become obese, their adipocytes enlarge, and adipose tissue suffers alterations at the molecular and cellular level affecting systemic metabolism. First, macrophages accumulate inside adipose tissue, which causes local inflammation, but also macrophages act as scavengers of apoptotic adipocytes^[28,32,58,61]. (**Figure 2A**)

Several pro-inflammatory factors are produced in adipose tissue (AT) of obese individuals in comparison with AT of lean individuals. Several studies have shown an increase in the secretion of cytokines like TNF- α and IL-6 that stimulate the preadipocytes and the endothelial cells to produce chemo-tactic (MCP-1). These can attract macrophages to the adipose tissue for induction of the inhibitor of titular plasminogen (PA1-1) and some adhesion molecules (P-selectin, ICAM, VCAM-1). It promotes adhesion, migration, and accumulation of monocytes and T lymphocytes in the subendothelial space as a consequence of the impairment in vascular permeability^[50]. Autophagy plays a fundamental role in inflammation, which is part of the obesity profile, influences the development, home-

Figure 2A.

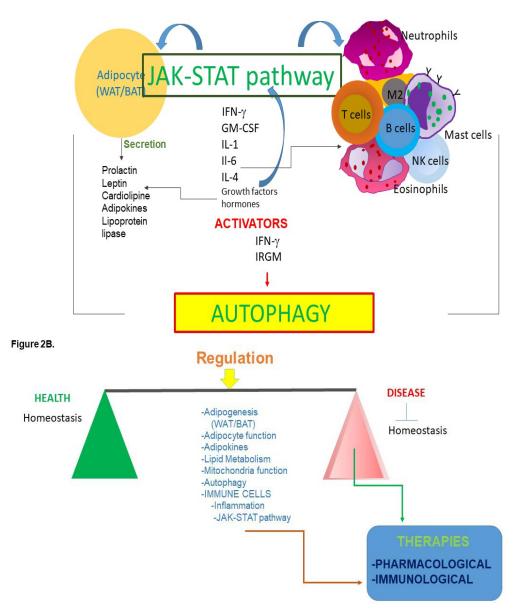


Figure 2. (**A**) The crosstalk in adipose tissue for modulating adipogenesis, the development, differentiation and the physiological function of adipocytes (white, beige-brown), involves to the immune cells and the cytokines produced, the autophagy machinery and the different secreted components by adipocytes. All of them behave as activators of the JAK-STAT pathway, that influences strongly the status of inflammation, considered as key endocrine immunological factor that trigger immunometabolism diseases. (**B**) The balance in the crosstalk and regulation of the physiological processes in adipose tissue with bias toward health or disease.

ostasis, and survival of the inflammatory cells, including macrophages, neutrophils, and lymphocytes, carried out transcription, processing, and secretion of a series of cytokines, besides to be regulated by other cytokines. It has shown that either IL-1 α or IL-1 β induce autophagy, which can behave as a negative feedback loop to control the inflammation induction by IL-1. The secretion of II-18 and TNF- α is regulated by autophagy. The inhibition of autophagy is due to the production of IL-18, reducing the production of IL-6, IL-8 and TNF- α ^[25,28,32,47]. Indeed, autophagy affect the secretion of cytokines type-TH1 (IFN- γ , TNF α , IL-1, IL-2, IL-6, TGF- β , MCP-1) and of the type-TH2 (IL-4, IL-10 and IL-13) as well as another type cytokines (IL-1 β , IL-18, IFN- α , IFN- β and IL-8)^[25,28]. The family of STATs or signal transducer activator of transcription (STATS) is activated by phosphorylation of one tyrosine (Tyr) residue near the C-terminus that is catalyzed by a Janus Kinase (JAK), conforming to the JAK-STAT signalization pathway. The STAT family of transcriptional activation play a pivotal role in transferring the external stimulus to the nucleus for expressing transcription factor that influences and participates in different immunological processes and in connection with endocrine system in adipogenesis development, differentiation and regulation. STAT1-3, STAT5, and STAT6 play a role directly or indirectly in human. For example, STAT1-3 participates through IFN-y modulation of adipocyte function, while STAT5 participates in the regulation of the differentiation of preadipocytes, adipocytes, myeloid and macrophage in adipose tissue^[21]. Another component of the innate immune cells, the GM-CSF (granulocyte-macrophage colony-stimulating factor) functions as a bridge between the innate and adaptive immune response. In normal conditions, GM-CSF secreted by tissue-invading lymphocytes plays a role in immunopathogenesis^[21,62]. (Figures 2A and 2B)

The level of the immunological response in adipose tissue refers to obesity. A series of biochemical reactions that are pivotal in the translation of positive or negative signals are the JAK-STAT signalization pathway. This pathway, the Janus kinase activation pathway (non-receptor tyrosine kinases) (JAK1, JAK2, JAK3, and TYK2)^[21,51], is formed by a diversity of ligand-, mediated signals, from cytokines and hormones, leads to activation downstream signaling pathways and alterations in gene expression. One of the physiological functions of the JKA activation is the immune effector function. Aberrant activation of JKA signaling plays a critical role in various chronic diseases, such as autoimmune disorders, several malignancies, and rheumatoid arthritis. The inhibitor of the JKA pathway is proposed as a new therapeutic approach^[24,63]. The JAK-STAT signalization pathway has a critical role in the crosstalk of adipocytes and immune cells in adipose tissue that might influence obesity pathogenicity. The family of JAKs (1-3) includes Ty2 and STATs (1-6) and is well known as critical in adipocyte differentiation (WAT/BAT) in AT. The immune cells produce pro-inflammatory and anti-inflammatory cytokines signals via the JAK-STAT pathway^[21,51]. In other words, the immune cells present in the adipose tissue participate dynamically and modulate the physiological function of either WAT or BAT adipocytes. Target genes of STAT pathways in AT have shed light on the role of the transcription factors in immunometabolism (adipocyte browning and whitening, insulin sensibility, lipid storage, and glucose homeostasis)^[58]. (**Figures 2A** and **2B**)

For example, STAT-1 transcriptionally regulates the lipoprotein lipase produced by adipocytes. Indeed, STATs activators play a role in the regulation of adipocyte differentiation. Moreover, these activators exhibit differential expression in conditions of obesity and/or insulin resistance. GM-CSF plays a role in adipose tissue to recruit and activate macrophages that contribute to AT inflammation and insulin resistance. Furthermore, the pro-inflammatory properties of IFN-y signaling through the JAK-STAT pathways can inhibit pre-adipocytes differentiation^[59,60,62,64]. In addition, IFN-g produced from infiltrated immune cells on adjacent adjpocytes leads to insulin resistance. It was observed in transgenic mice in the IFN- γ gene that the shift from ATM in WAT toward a phenotype BAT in ATMs results in decreased production of inflammatory cytokines and improved insulin sensitivity^[28,32,52,63,65]. (Figures 2A and 2B)

4. Conclusion

For immunometabolism disease therapeutic interventions (i.e. obesity), it makes necessary to take advantage of the crosstalk of the endocrine, immunological, and autophagy programs. One alternative could be to promote the browning of white adipose tissue, possibly through keeping or modulating the inflammation in AT macrophage activation (IL-6 production) and species reactive oxygen, inhibiting fatty acid release but promoting fatty-acid combustion (exercise). A second alternative is to target inflammation modulation by innate cells in AT through the JAK-STAT signalization pathway with the interplay with the autophagy machinery. For example, ATG proteins are associated with autophagy machinery targeted by Chloroquine treatment, and this might influence the outcome of the endocrine-immunological response potentially in AT of obese individuals.

A third one is, to target autophagy, by subtle

inhibition through pharmacological drugs) which will impact adipocyte differentiation. Moreover, to inhibit or modulate autophagy expression proteins (ATG-5), and downregulate IFN- γ production and TLRs expression on macrophages. By another hand, the modulation of ATMs in AT and the products secreted by them is pivotal for insulin sensitivity regulation.

An exciting area of basic and clinical research is the translation of the crosstalk and the status of the adipocytes, immune cells, and autophagy. The three have the enormous task of favoring steady low inflammation and adipocyte browning to get healthier tissues and longer life.

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Conflict of interest

No conflict of interest was declared by the author.

References

- Caballero B. Human against obesity. Who will win? Advances in Nutrition 2019; 10(suppl_1): S4–S9. doi: 10.1093/advances/nmy055
- 2. WHO. Obesity and overweight [Internet]. 2021. Available from: https://www.WHO.Int/es/news-room/fact-sheets/det ail/obesity-and-overweight
- Gutin B. Child obesity can be reduced with vigorous activity rather than restriction of energy intake. Obesity 2008; 16(10): 2193–2196. doi: 10.1038/oby.2008.348
- 4. Hill JO, Wyatt HR, Peters JC. Energy balance and obesity. Circulation 2012; 126(1): 126–132. doi: 10.1161/CIRCULATIONAHA.111.087213
- Howell S, Kones R. "Calories in, calories out" and macronutrient intake: The hope, hype, and science of calories. American Journal of Physiology-Endocrinology and Metabolism 2017; 313(5): E608–E612. doi: 10.1152/ajpendo.00156.2017
- Namboong S, Cho Ch-S, Semple I, Lee JH. Autophagy dysregulation and obesity-associated pathologies. Molecules and Cells 2018; 41(1): 3–10. doi: 10.14348/molcells.2018.2213
- Shao F, Chen Y, Xu H, *et al.* Metabolic obesity phenotypes and risk of lung cancer: A Prospective cohort study of 450,482 UK biobank participants. Nutrients 2022; 14(16): 3370. doi: 10.3390/nu14163370

- Shinjyo N, Kita K. Infection and immunometabolism in the central nervous system: A possible mechanistic link between metabolic imbalance and dementia. Frontiers in Cellular Neuroscience 2021; 15: 765217. doi: 10.3389/fncel.2021.765217
- Flores-Cordero JA, Pérez-Pérez A, Jiménez-Cortegana C, *et al.* Obesity as a risk factor for dementia and Alzheimer's disease: The role of leptin. International Journal of Molecular Sciences 2022; 23(9): 5202. doi: 10.3390/ijms23095202
- Liu J, Zhen D, Hu C, *et al.* Reconfiguration of gut microbiota and reprogramming of liver metabolism with phycobiliproteins bioactive peptides to rehabilitate obese rats. Nutrients 2022; 14(17): 3635. doi: 10.3390/nu14173635
- Wen X, Zhang B, Wu B, *et al.* Signaling pathways in obesity: Mechanisms and therapeutic interventions. Signal Transduction and Targeted Therapy 2022; 7(1): 298. doi: 10.1038/s41392-022-01149-x
- Ghaben AI, Scherer PE. Adipogenesis and metabolic health. Nature Reviews Molecular Cell Biology 2019; 20: 242–258. doi: 10.1038/s41580-018-0093-z
- Haider N, Larose L. Harnessing adipogenesis to prevent obesity. Adipocyte 2019; 8(1): 98–104. doi: 10.1080/21623945.2019.1583037
- Scheele C, Wofrum Ch. Brown adipose crosstalk in tissue plasticity and human metabolism. Endocrine Reviews 2020; 41: 53–65. doi: 10.1210/endrev/bnz007
- 15. Haczeyni F, Bell-Anderson KS, Farrell GC. Cause and mechanism of adipocyte enlargement and adipose expansion. Obesity Reviews 2018; 19(3): 406– 420. doi: 10.1111/obr.12646
- Longo M, Zatterale F, Naderi J, *et al.* Adipose tissue dysfunction as determinant of obesity-associated metabolic complications. International Journal of Molecular Sciences 2019; 20(9): 2358. doi: 10.3390/ijms20092358
- Jia L, Chen Z, Pan T, *et al.* TRIM67 deficiency exacerbates hypothalamic inflammation and fat accumulation in obese mice. International Journal of Molecular Sciences 2022; 23(16): 9438. doi: 10.3390/ijms23169438
- Ouchi N, Parker JI, Lugus JJ, Walsh K. Adipokines in inflammation and metabolic disease. Nature Reviews Immunology 2011; 11(2): 85–97. doi: 10.1038/nri2921
- Singla P, Bardoloi A, Parkash AA. Metabolic effects of obesity: A review. World Journal of Diabetes 2010; 1(3): 76–88. doi: 10.4239/wjd.v1.i3.76
- 20. Caspar-Bauguil S, Cousin B, Bour S, *et al.* Adipose tissue lymphocytes and roles. Journal of Physiology and Biochemistry 2009; 65: 423–436. doi: 10.1007/BF03185938
- Richard AJ, Stephens JM. The role of JAK-STAT signaling in adipose tissue function. Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease 2014; 1842(3): 431–439. doi: 10.1016/j.bbadis.2013.05.030

- Matsuzawa Y. Adiponectin: A key player in obesity related disorders. Current Pharmaceutical Design 2010; 16(17): 1896–1901. doi: 10.2174/138161210791208893
- Yamawaki H, Kuramoto J, Kameshima S, *et al.* Omentin, a novel adipocytokine inhibits TNF-induced vascular inflammation in human endotelial cells. Biochemical and Biophysical Research Communications 2011; 408: 339–343. doi: 10.1016/j.bbrc.2011.04.039
- AL-Suhaimi AE, Shehzad A. Leptin, resistin, and visfatin: The missing link between endocrine metabolic disorders and immunity. European Journal of Medical Research 2013; 18(1): 12. doi: 10.1186/2047-783X-18-12
- Kern L, Mittenbühler MJ, Vesting AJ, *et al.* Obesity-induced TNFα and IL-6 signaling: The missing link between obesity and inflammation-driven liver and colorectal cancers. Cancers 2018; 11(1): 24. doi: 10.3390/cancers11010024
- Freff J, Schwarte K, Bröker L, *et al.* Alterations in B cell subsets correlate with body composition parameters in female adolescents with anorexia nervosa. Scientific Reports 2021; 11(1): 1125. doi: 10.1038/s41598-020-80693-4
- Herz CT, Kiefer FW. Adipose tissue browning in mice and humans. Journal of Endocrinology 2019; 241(3): R97–R109. doi: 10.1530/JOE.18-0598
- Ingelfinger F, De Feo D, Becher B. GM-CSF: Master regulator of the T cell-phagocyte interface during inflammation. Seminars in Immunology 2021; 54: 101518. doi: 10.1016/j.smim.2021.101518
- 29. Ro S-H, Jang Y, Bae J, *et al.* Autophagy in adipocyte browning: Emerging drug target for intervention in obesity. Frontiers in Physiology 2019; 10: 22. doi: 10.3389/fphys.2019.00022
- 30. Liu GY, Sabatini DM. mTOR at the nexus of nutrition, growth, aging, and disease. Nature Reviews Molecular Cell Biology 2020; 21(4): 183–203. doi: 10.1038/s41580-019-0199-y
- Morishita H, Mizushima N. Diverse cellular roles of autophagy. Annual Review of Cell and Developmental Biology 2019; 35: 3.1–3.23. doi: 10.1146/annual-cellbio-100818-125300
- 32. Deretic V. Autophagy in inflammation, infection, and immunometabolism. Immunity 2021; 54(3): 437–453. doi: 10.1016/j.immuni.2021.01.018
- 33. Wang J, Liao B, Wang C, *et al.* Effects of antioxidant supplementation on metabolic disorders in obese patients from randomized clinical controls: A meta-analysis and systematic review. Oxidative Medicine and Cellular Longevity 2022; 2022: 7255413. doi: 10.1155/2022/7255413
- Zhang Y, Goldman S, Baerga R, *et al.* Adipose-specific deletion of autophagy-related gene 7 (atg7) in mice reveals a role in adipogenesis. Proceedings of the National Academy of Sciences 2009; 106(47): 19860–19865. doi: 10.1073/pnas.0906048106
- 35. Quan W, Lee MS. Role of autophagy in the control

of body metabolism. Endocrinology and Metabolism 2013; 28(1): 6–11. doi: 10.3803/EnM.2013.28.1.6

- Kovsan J, Bluher M, Tarnovscki T, *et al.* Altered autophagy in human adipose tissues in Obesity. The Journal of Clinical Endocrinology and Metabolism 2011; 96(2): E268–E277. doi: 10.1210/jc.2010-1681
- 37. Boya P, Reggiori F, Codogno P. Emerging regulation and functions of autophagy. Nature Cell Biology 2013; 15(7): 713–720. doi: 10.1038/ncb2788
- Zheng ZG, Zhu ST, Cheng HM, *et al.* Discovery of a potent SCAP degrader that ameliorates HFD-induced obesity, hyperlipidemia and insulin resistance via an autophagy-independent lysosomal pathway. Autophagy 2021; 17(7): 1592–1613. doi: 10.1080/15548627.2020.1757955
- Menikdiwela KR, Ramalingam L, Rasha F, *et al.* Autophagy in metabolic syndrome: Breaking the wheel by targeting the renin-angiotensin system. Cell Death & Disease 2020; 11: 87–94. doi: 10.1038/s41419-020-2275
- 40. Park HS, Song JW, Park JH, *et al.* TXNIP/VDUP1 attenuates steatohepatitis via autophagy and fatty acid oxidation. Autophagy 2021; 17(9): 2549–2564. doi: 10.1080/15548627.2020.1834711
- 41. Yang H, Wen Y, Zhang M, *et al.* MTORC1 coordinates the autophagy and apoptosis signaling in articular chondrocytes in osteoarthritic temporomandibular joint. Autophagy 2020; 16(2): 271–288. doi: 10.1080/15548627.2019.1606647
- 42. Karampela I, Christodoulatos GS, Dalamaga M. The role of adipose tissue and adipokines in sepsis: In-flammatory and metabolic considerations, and the obesity paradox. Current Obesity Reports 2019; 8(4): 434–457. doi: 10.1007/s13679-019-00360-2
- 43. Mizushima N. Autophagy: Process and function. Genes & Development 2017; 21(22): 2861–2873.
- Cristancho AG, Lazar MA. Forming functional fat: A growing understanding of adipocyte differentiation. Nature Reviews Molecular Cell Biology 2011; 12: 722–734. doi: 10.1038/nrm3198
- Sarjeant K, Stephens JM. Adipogenesis. Cold Spring Harbor Perspectives in Biology 2012; 4: a008417– a008436.
- 46. Chun Y, Kim K. Autophagy: An essential degradation program for cellular homeostasis and life. Cells 2018; 7(12): 278–304. doi: 10.3390/cells7120278
- 47. Qian M, Fang X, Wang X. Autophagy and inflammation. Clinical and Translational Medicine 2017; 6(1): e24. doi: 10.1186/s40169-017-0154-5
- Yuan ML, Wang T. The new mechanism of Ghrelin/GHSR-1a on autophagy regulation. Peptides 2020; 126: 170264. doi: 10.1016/j.peptides.2020.170264
- 49. Tao T, Xu H. Autophagy and obesity and diabetes. In: Le W (editor). Autophagy: Biology and diseases. Advances in experimental medicine and biology, vol 1207. Singapore: Springer; 2020. p. 445–461. doi: 10.1007/978-981-15-4272-5_32
- 50. Delgado MA, Elmaoued RA, David AS, *et al.* Toll-like receptors control autophagy. The EMBO

Journal 2008; 27(7): 1110–1121. doi: 10.1038/emboj.2008.31

- Levine RI, Hubbard SR. Unlocking the secrets to Janus kinase activation. Science 2022; 376(6589): 139–140. doi: 10.1126/Science.Abo7788
- 52. Deretic V, Levine B. Autophagy balance inflammation in innate immunity. Autophagy 2018; 14: 243– 251.
- Hu Y, Reggiori F. Molecular regulation of autophagosome formation. Biochemical Society Transactions 2022; 50(1): 55–69. doi: 10.1042/BST20210819.
- Mizushima N, Levine B. Autophagy in human diseases. New England Journal of Medicine 2020; 383(16): 1564–1576. doi: 10.1056/NEJMra2022774
- Zhao YG, Zhang H. Formation and maturation of autophagosomes in higher eukaryotes: A social network. Current Opinion in Cell Biology 2018; 53: 29–36. doi: 10.1016/j.ceb.2018.04.003
- Wang Z, Nakayama T. Inflammation, a link between obesity and cardiovascular disease. Mediators of Inflammation 2010; 2010: 535918. doi: 10.1155/2010/535918
- Singh R, Xiang Y, Wang Y, *et al.* Autophagy regulates adipose mass and differentiation in mice. The Journal of Clinical Investigation 2009; 119(11): 3329–3339. doi: 10.1172/JCI39228
- Farkhondeh T, Llorens S, Pourbagher-Shahri AM, *et al.* An overview of the role of adipokines in cardiometabolic diseases. Molecules 2020; 25(21): 5218. doi: 10.3390/molecules25215218

- 59. Miller BC, Zhan Z, Stephenson LM, *et al.* The autophagy gene ATG5 plays an essential role in B lymphocyte development. Autophagy 2008; 4(3): 309–314. doi: 10.4161/auto.5474
- Clarke AJ, Simon AK. Autophagy in the renewal differentiation and homeostasis of immune cells. Nature Reviews Immunology 2019; 19(3): 170–183. doi: 10.1038/s41577-018-0095-2
- 61. Mills EL, Kelly B, O'Neill LAJ. Mitochondria are the powerhouses of immunity. Nature Immunology 2017; 18; 488–498. doi: 10.1038/ni.3704
- 62. Villarino A, Laurence A, Robinson GW, *et al.* Signal transducer and activator of transcription 5 (STAT5) paralog dose governs T cell effector and regulatory functions. eLife 2016; 5: e08384. doi: 10.7554/eLife.08384
- Gonciarz M, Pawlak-Bus, Leszczynski P, Owczarek W. TYK2 as a therapeutic target in the treatment of autoimmune and inflammatory diseases. Immunotherapy 2021; 13(13): 1135–1150. doi: 10.2217/imt-2021-0096
- Kumar S, Jain A, Choi SW, *et al.* Mammalian Atg8-family proteins are upstream regulators of the lysosomal system by controlling mTOR and TFEB. Autophagy 2020; 16: 2305–2306. doi: 10.1080/15548627.2020.1837423
- 65. Khan IM, Dai Perrard XY, Perrard JL, *et al.* Attenuated adipose tissue and skeletal muscle inflammation in obese mice with combined CD4+ and CD8+ T cell deficiency. Atherosclerosis 2014; 233(2): 419–428. doi: 10.1016/j.atherosclerosis.2014.01.011